

^a Reagents: (i) 1:1 dioxane-H₂O, K₂CO₃, BOC anhydride, 25 °C, 3.5 h; (ii) MeOH, PhCHN₂, 25 °C, 72%; (iii) TFA, EtOAc, 0 °C to 25 °C, 30 min, evaporate; 2 equiv of N-methylmorpholine, DMF, 16, 25 °C, 72 h, 53%; (iv) TBAF, THF, 25 °C, 30 min; (v) 10% Pd/BaSO₄ (prehydrogenated overnight), MeOH/H₂O, H₂, 25 °C, 30 min, 39%.

Although saponification of the ester 13 led to undesired desilylation and lactonization, ester 13 could be hydrolyzed under nonaqueous conditions developed in this laboratory.²² Transesterification of 13 in the presence of 3-buten-1-ol and Ti(Oi-Pr)₄²³ gave the ester 14 (91%), which was protected as the *tert*-butyldimethylsilyl ether 15 (95%).²⁴ Ozonolysis of the butene residue afforded the aldehyde which, after treatment with DBU²² in situ, gave the α -azido acid. Without isolation, treatment of the acid with *p*-nitrophenol and DCC in THF²⁵ afforded the activated ester 16 (61%) as a single diastereoisomer.

p-Nitrophenyl esters are widely used to facilitate peptide bond formation.^{25,26} In fact, a variety of *p*-nitrophenyl esters of N-protected α -amino acids have been utilized in the peptide-coupling reaction with α -amino nucleosides

Principles of Peptide Synthesis; Springer, Verlag: New York, 1984; p 30.

including the C-terminal amino acid, uracil polyoxin C (4).²⁷ Unfortunately, reaction of the C-terminal amino acid 4 with the activated ester 16 in DMF^{27b} failed to afford any coupled product. However, after protecting nucleoside 4, formation of the amide bond was achieved. The α -amino acid was first protected as the BOC derivative²⁸ followed by formation of the benzyl ester 17 using PhCHN₂²⁹ (72% over 2 steps) (Scheme III). Liberation of the amine moiety by treatment with trifluoroacetic acid²⁶ gave the corresponding ammonium trifluoroacetate which was used directly in the peptide coupling reaction in DMF using N-methylmorpholine,²⁷ affording amide 18 (53%). Desilylation¹⁷ of 18 and subsequent chemoselective hydrogenation over 10% Pd/BaSO₄ in aqueous methanol^{11c} afforded the nucleoside dipeptide amino acid nikkomycin B (1) $(39\%).^{30}$

In conclusion, a potentially general route to the (2S,3S,4S)- γ -hydroxy- β -methyl- α -aminobutanoic acid moiety, a common feature of the nikkomycin N-terminal amino acid residues, has been described. The protected α -amino nucleoside fragment 17 has been successfully coupled with the activated *p*-nitrophenyl ester 16, completing the total synthesis of nikkomycin B (1).

Acknowledgment. We thank the National Institutes of Health (AI-22252) for the support of this program and for the purchase of a 400-MHz NMR spectrometer (RR-01672) and a high-resolution mass spectrometer (RR-03245) used in these studies. We additionally thank Dr. Colin Smith of Glaxo Research Group, Greenford, Middlesex, U.K., for most generously providing authentic uracil polyoxin C (4) for studies on the peptide coupling reaction, G. D. Searle and Co., Skokie, IL, for microanalytical services, and Dr. Xiao-an Zhang for preliminary studies.

Supplementary Material Available: Experimental procedure for the conversion of 18 into nikkomycin B (1) and spectroscopic data for 1 (2 pages). Ordering information is given on any current masthead page.

(29) Adamson, J. R.; Bywood, R.; Eastlick, D. T.; Gallagher, R.; Walker, D.; Wilson, E. M. J. Chem. Soc., Perkin Trans. 1 1975, 2030.

(30) We have been unable to obtain an authentic sample of nikkomycin B (1). Our structural assignment is based upon comparison of spectroscopic data for our synthetic material with the data reported by Konig for nikkomycin B_x (2). See ref 6 and the supplementary material.

Short Synthesis of (\pm) -Sterpurene

Shi-Kai Zhao and Paul Helquist*

Department of Chemistry and Biochemistry, University of Notre Dame, Notre Dame, Indiana 46556 Received September 11, 1990

Summary: The tricyclic fungal metabolite sterpurene (1) has been synthesized by a very short route employing a cyclopentane annulation as the key transformation.

Sterpurene (1) was first reported by Ayer in 1981 as a metabolite of *Stereum purpureum*, a fungus that is responsible for silver leaf disease.¹ Syntheses of racemic and

nonracemic sterpurene have been reported by Murata, Little, and Okamura.² Herein we report our synthesis of (\pm) -sterpurene.

⁽²²⁾ Barrett, A. G. M.; Lebold, S. A.; Zhang, X.-A. Tetrahedron Lett. 1989, 30, 7317.

⁽²³⁾ Seebach, D.; Hungerbühler, E.; Naef, R.; Schnurrenberger, P.;
Weidmann, B.; Züger, M. Synthesis 1982, 138.
(24) Corey, E. J.; Cho, H.; Rücker, C.; Hua, D. H. Tetrahedron Lett.

⁽²⁴⁾ Corey, E. J.; Cho, H.; Rücker, C.; Hua, D. H. Tetrahedron Lett. 1981, 22, 3455.

 ⁽²⁵⁾ Bodansky, M.; du Vigneaud, V. J. Am. Chem. Soc. 1959, 81, 5688.
 (26) (a) Bodansky, M.; Klausner, Y. S.; Ondetti, M. A. Peptide Synthesis; John Wiley: New York, 1976; pp 102-106. (b) Bodansky, M.

 ^{(27) (}a) Azuma, T.; Saita, T.; Isono, K. Chem. Pharm. Bull. 1977, 25, 1740.
 (b) Khare, R. K.; Becker, J. M.; Naider, F. R. J. Med. Chem. 1988, 31, 650.

⁽²⁸⁾ Moroder, L.; Hallett, A.; Wünsch, E.; Keller, O.; Wersin, G. Hoppe-Seyler's Z. Physiol. Chem. 1976, 357, 1651.

⁽¹⁾ Ayer, W. A.; Saeedi-Ghomi, M. H. Can. J. Chem. 1981, 59, 2536.

^{(2) (}a) Murata, Y.; Ohtsuka, T.; Shirahama, H.; Matsumoto, T. Tetrahedron Lett. 1981, 22, 4313. (b) Moëns, L.; Baizer, M. M.; Little, R. D. J. Org. Chem. 1986, 51, 4497. (c) Gibbs, R. A.; Okamura, W. H. J. Am. Chem. Soc. 1988, 110, 4062. (d) Gibbs, R. A.; Bartels, K.; Lee, R. W. K.; Okamura, W. H. Ibid. 1989, 111, 3717.

5821



cis-6-Methylbicyclo[4.2.0]oct-3-en-2-one (2)³ undergoes copper-catalyzed conjugate addition⁴ of isobutylmagnesium bromide to give the ketone 3 as a 4:1 mixture of the β and α isomers favoring the desired β -substituted isomer. Enolate generation followed by addition of $[Cp(CO)_2Fe=$ CHSPh]⁺PF₆⁻ (5, Cp = η^5 -C₅H₅)⁵ gives the intermediate 6 which when treated with trimethyloxonium tetrafluoroborate gives the epimeric tricyclic ketones 7⁶ (Scheme I). This product is obtained in 80–90% yields from 6, but the overall cyclopentane annulation is made operationally simpler when 6 is not isolated or purified, in which case the overall yield of 7 is 48% from the enol silyl ether 4. The main limitation in the yield for the overall annulation process appears to be incomplete reaction of the enolate generated from 4 in that 20-30% of the ketone 3 is recovered after the reaction of the enolate with 5. The synthesis is completed as in Little's route^{2b} by addition of methyllithium and dehydration of the resulting tertiary alcohol. The sterpurene thus obtained is identical to the previously obtained material according to direct comparison of ¹H and ¹³C NMR spectra.^{1,2d}

This synthesis provides sterpurene by an attractively simple, short route, and it demonstrates the utility of the new organoiron-based cyclization reaction.⁶

Acknowledgment. We thank Professors Ayer and Okamura for generously providing us with copies of NMR spectra of sterpurene. We also thank Dr. Bruce Plashko (Notre Dame) for mass spectrometric measurements. We are grateful to the National Science Foundation and the donors of the Petroleum Research Fund, administered by the American Chemical Society, for financial support of this work.

Supplementary Material Available: ¹H and ¹³C NMR spectra of *trans*-7 and the synthetic sterpurene (8 pages). Ordering information is given on any current masthead page.

Addition of Amino Lactams to Vinyl Vicinal Tricarbonyls. Formation of Tricyclic 2-Azadethiapenams and 3-Azadethiacephams

Harry H. Wasserman,* Susan L. Henke, Patrick Luce, and Eiji Nakanishi

Department of Chemistry, Yale University, New Haven, Connecticut 06511

Gayle Schulte

Yale Instrument Center, Yale University, New Haven, Connecticut 06511 Received August 29, 1990

Summary: Amino lactams react as trinucleophiles with vinyl vicinal tricarbonyl esters to form tricyclic derivatives. Products of particular interest resulting from this reaction sequence are 2-azadethiapenams and 3-azadethiacephams.

We have recently shown¹ that the addition of primary amines to vinyl tricarbonyl reagents A generates ketopyrrolinium carboxylates B (Scheme I), which may serve as powerful electrophilic acceptors for nucleophilic residues attached to the primary amine (Figure 1). These nucleophiles have included indole groups, activated aromatic rings, enol ethers, vinylsilanes, propargylsilanes, and pyrroles.¹⁻³ We now report that the amide NH of a lactam



residue will add to the intermediate iminium group, forming fused-ring 2-(acylamino)-1-oxopyrrolidinecarboxylates.⁴ This process constitutes a facile route to

⁽³⁾ Enone 2 was obtained as previously reported from the photocycloaddition of ethylene to 3-methyl-2-cyclohexenone followed by bromination/dehydrobromination: (a) Zurflüh, R.; Dunham, L. L.; Spain, V. L.; Siddall, J. B. J. Am. Chem. Soc. 1970, 92, 425. (b) Cargill, R. L.; Dalton, J. R.; Morton, G. H.; Caldwell, W. E. Org. Synth. 1984, 62, 118. (c) Jacques, J.; Marquet, A. Organic Syntheses; Wiley: New York, 1988; Collect. Vol. VI, p 175. (d) House, H. O.; Bashe, R. W. II J. Org. Chem. 1965, 30, 2942.

^{(4) (}a) Horiguchi, Y.; Matsuzawa, S.; Nakamura, E.; Kuwajima, I. Tetrahedron Lett. 1986, 27, 4025. (b) Bal, S. A.; Marfat, A.; Helquist, P. J. Org. Chem. 1982, 47, 5045. Direct trapping of the enolate resulting from the conjugate addition was performed using TMSCl, and direct trapping of the enolate resulting from proton abstraction from ketone 3 was performed using iron reagent 5, but superior results were obtained with the more stepwise route described in the text.

^{(5) (}a) Knors, C.; Kuo, G.-H.; Lauher, J. W.; Eigenbrot, C.; Helquist, P. Organometallics 1987, 6, 988. (b) Knors, C.; Helquist, P. Organomet. Synth. 1988, 4, 205.

⁽⁶⁾ Zhao, S.-K.; Knors, C.; Helquist, P. J. Am. Chem. Soc. 1989, 111, 8527.

⁽¹⁾ Wasserman, H. H.; Fukuyama, J.; Murugesan, N.; van Duzer, J.; Lombardo, L.; Rotello, V.; McCarthy, K. J. Am. Chem. Soc. 1989, 111, 371-372.

⁽²⁾ Wasserman, H. H.; Amici, R. M. J. Org. Chem. 1989, 54, 5843–5844.
(3) Wasserman, H. H.; Cook, J. D.; Vu, C. B. J. Org. Chem. 1990, 55, 1701–1702.